

Review Article

Biological applications of biosynthesized silver nanoparticles through the utilization of plant extracts

Rouhollah Heydari*

Razi Herbal Medicines Research Center, Lorestan University of Medical Sciences, Khorramabad, Iran

Received: 08.05.2017; Accepted: 21.06.2017

Abstract

Widespread uses of metallic nanoparticles, especially silver nanoparticles (AgNPs) in biology, pharmaceuticals, and medicine lead to the development of biosynthesis methods that are in turn utilized to prepare these nanoparticles. Among the biosynthesis methods, which are used to prepare nanoparticles, the plant-mediated methods have gained great attention due to several advantages such as cost-effectiveness, availability, eco-friendliness and nontoxicity of plants. Moreover, plant extracts are rich in different compounds which act as inhibitory and capping agents. For these reasons, plant-mediated methods can be potentially used for large-scale production of nanoparticles with different properties. The present article focuses on plant-mediated AgNPs using various plants and their biological applications such as antimicrobial, antioxidant, anticancer, anti-inflammatory, hepatoprotective and antilarvicidal properties.

Keywords: Silver nanoparticles, Plant extract, Antimicrobial, Antioxidant, Anticancer

***Corresponding Author:** Dr. Rouhollah Heydari, Razi Herbal Medicines Research Center, Lorestan University of Medical Sciences, Khorramabad, Iran. Email: rouhollahheydari@yahoo.com.

Please cite this article as: Heydari R. Biological applications of biosynthesized silver nanoparticles through the utilization of plant extracts. *Herb. Med. J.* 2017;2(2):87-95.

Introduction

Over the last few years, a variety of inorganic nanomaterials such as nanoparticles, nanowires, and nanotubes have been created or modified in order to obtain superior properties with greater functional versatility. The nanoscale science and technology significantly contribute to the development of new strategies for the synthesis of uniform nanomaterials with the controlled size and shape. In particular, nanoparticles in the range of 1–100 nm, have been investigated due to their size for their uses as tools for a new generation of technological devices. Moreover, due to the similarity of their dimensions

and shapes to several biological structures (e.g., membrane cell genes, proteins, and viruses), they have been proposed for investigating biological processes as well as for sensing and treating diseases (1).

Many studies have focused on silver nanoparticles (AgNPs) due to their wide variety of applications (2). The AgNPs have widespread biological activities such as antimicrobial (3), anthelmintic (4), antilarvicidal (5), antioxidant (6), anticancer (7), anti-inflammatory (8), hepatoprotective (9), and wound healing activities (10).

The review of literature in the time span between 1987 to 2017 using silver nanoparticles as a keyword in "Scopus" database resulted in 23,312 articles (Figure

1) from which 3,169 articles deal with medicinal issues (Figure 2). Furthermore, the search results were refined by several keywords such as plant extracts, biosynthesis, green synthesis and biological applications which resulted in 1797, 2428, 5705 and 7302 articles, respectively.

Green chemistry refers to the activity of designing chemical products and processes that reduce or eliminate the use or generation of substances which are hazardous for human health and environment. Therefore, green chemistry protects the environment, not by cleaning up, but by introducing new chemical processes that do not pollute the environment. In this review, the green synthesis of AgNPs using plant extracts and their biological applications has been discussed.

Conventional Methods of AgNPs Synthesis

Several methods, such as chemical synthesis (11), electrochemical (12), radiation (13, 14), photochemical (15) and biological synthesis (16-19) have been used for the synthesis of AgNPs. Chemical, electrochemical, radiation and photochemical methods are expensive, energy consuming, harmful for both human beings and environment, and not suitable for biological applications. In comparison with these methods, biological methods used for synthesis of AgNPs are cost-effective, safe and environmental friendly. Consequently, these methods have been preferred as green chemistry methods.

Plant-mediated AgNPs Synthesis

Nowadays, the development of green synthesis of metallic nanoparticles and their applications are considered to be among the most important areas of research. Among the biological methods, the use of plant extracts for the synthesis of AgNPs is simple and cost-effective. Moreover, the synthesized particles are stable. On the other hand, plants possess secondary metabolites which lead to the reduction of metal nanoparticles in the easiest way (20). Recently, a rapid, energy-efficient, green and economically scalable room temperature method for the synthesis of stable AgNPs by the use of tannic acid (a polyphenolic compound derived from plant extract) was developed by Sivaraman et al (21). The proposed mechanism for biosynthesis of AgNPs has been illustrated in scheme 1.

Various plant species and their distinct components have been reported for the synthesis of AgNPs. The data indicated in Table 1 concern the plant species and that part of them which has been used, average size, shape and application of biosynthesized AgNPs. As it can be observed, most biosynthesized AgNPs are spherical in shape and have an average size less than 100 nm.

Applications of Biosynthesized AgNPs

Several reports demonstrated that the antimicrobial activities of AgNPs depend on the size, shape and stabilizing agents of nanoparticles. The antibacterial activities increase as the size of AgNPs is reduced (22, 23). Aggregation of nanoparticles reduces antibacterial activities of AgNPs. Therefore, combination of nanoparticles with stabilizer agents prevents the aggregation and leads to the maintenance of antibacterial activities of AgNPs.

Antimicrobial Properties

Shankar et al., used the *Rhomdomyrtus tomentosa* acetone extract (RAE) as reducing and capping agents for the synthesis of gold, silver and gold-silver-alloy nanoparticles. The obtained nanoparticles were in the range of 10-100 nm. Fourier-transform infrared spectroscopy (FT-IR) spectra demonstrated that compounds in RAE were capped on the nanoparticles. This phenomenon leads to the extension of the antibacterial activity of RAE towards Gram-negative bacteria (39).

The antibacterial activity of biosynthesized AgNPs using *Anredera cordifolia* extract was investigated against several Gram-positive strains (*Staphylococcus aureus* and *Bacillus cereus*) and Gram-negative strains (*Escherichia coli*, *Pseudomonas aeruginosa*, *Proteus vulgaris*, and *Klebsiella pneumonia*). The results showed that the inhibition zones for six organisms were as follow: *E. coli* (16.33±0.58 mm), *B. cereus* (15.33±0.58 mm), *S. aureus* (14.67±0.58 mm), *K. pneumonia* (13.67±0.58), *P. aeruginosa* (13.33±0.58), and *P. vulgaris* (13.00±0.00). The antibacterial activity of biosynthesized AgNPs from aqueous extract of *A. cordifolia* was compared with the standard drug, gentamicin, which indicated that the biosynthesized AgNPs have significant antibacterial activity against six organisms (42).

Another study indicated that the diameters of the inhibition zones of biosynthesized AgNPs using

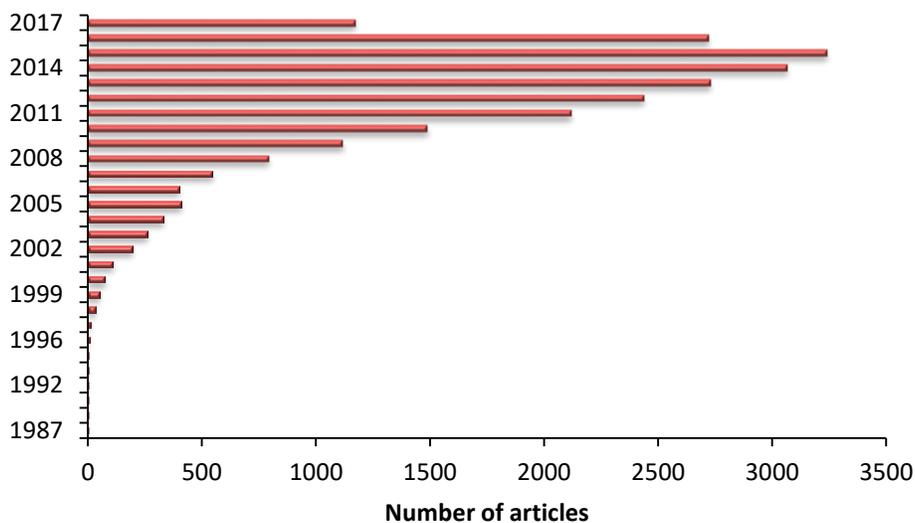


Figure 1. Number of articles in the time span between 1987 to 2017 using silver nanoparticles as keyword..

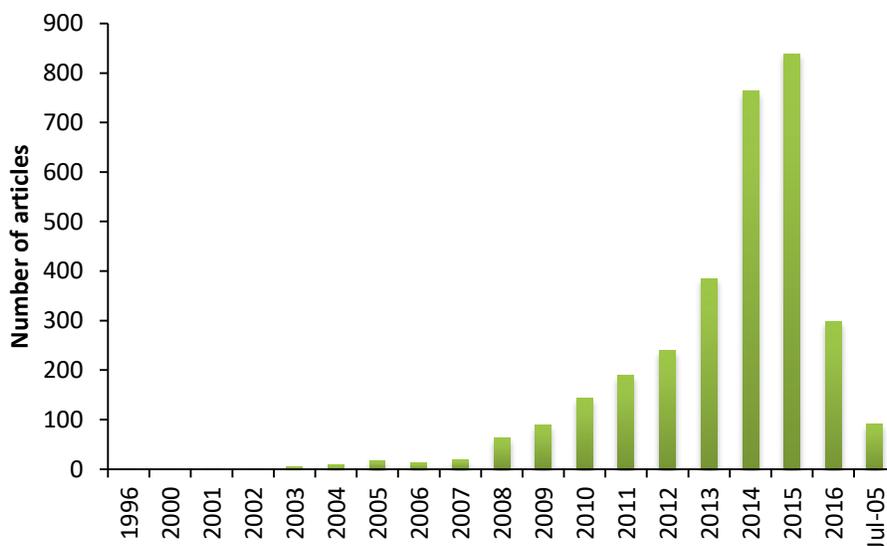


Figure 2. Number of articles dealing with medicinal topics in the time span between 1996 to 2017 using silver nanoparticles as keyword.

Tamarindus indica fruit extract against *B. cereus*, *S. aureus*, *Micrococcus luteus*, *Bacillus subtilis*, and those of *Enterococcus sp.* are 15, 16, 14, 18, and 16 mm respectively and *Pseudomonas aeruginosa*, *Salmonella typhi*, *E. coli*, and *K. pneumonia* are 22, 15, 15, and 10 mm respectively. In this study, the diameter of the inhibition zones of biosynthesized AgNPs were compared with AgNO₃. The results showed that the antibacterial activity of biosynthesized AgNPs against studied organisms was greater than that of AgNO₃ (43).

The biosynthesized AgNPs using corn leaf waste of *Zea mays* were evaluated for their antibacterial activity against foodborne pathogenic bacteria (*B. cereus*, *Listeria monocytogenes*, *S. aureus*, *E. coli* and *Salmonella Typhimurium*) by Patra et al., (46). The AgNPs at 50 µg/disk exhibited a moderate level of antibacterial activity against all five pathogenic bacteria with the diameter of inhibition zones of 9.26–11.57 mm. Moreover, synergistic antibacterial activity of biosynthesized AgNPs, together with the standard antibiotics, kanamycin and rifampicin, were examined

against the above-mentioned five foodborne pathogenic bacteria. The results demonstrated that the use of antibiotic and biosynthesized AgNPs together enhanced the antibacterial activity against all pathogens.

Rajakumar et al., reported that leaf extract of *Millettia pinnata* and biosynthesized AgNPs possess antibacterial potential against *E. coli*, *Pseudomonas aeruginosa*, *P. vulgaris*, *S. aureus* and *K. pneumonia*. This study suggests that the antibacterial activity of AgNPs could be fulfilled in three mechanisms (49). First, the AgNPs could adhere to bacterial cell wall and deactivate the cellular enzyme due to their fine size and large surface area. Therefore, permeability of the cell membrane of bacteria increased which in turn led to cell death (51). Second, interactions of AgNPs with the thiol group of L-cysteine protein may lead to enzymatic dysfunction (52). Finally, the AgNPs may facilitate the release of reactive oxygen species (ROS) which lead to cell death (53).

Antioxidant Activity

The antioxidant activity of biosynthesized AgNPs using *Anredera cordifolia* leaf extract was evaluated using 2,2-diphenyl-1-picrylhydrazyl (DPPH). In comparison with gallic acid as a standard case, antioxidant activity of biosynthesized AgNPs exhibited an effective inhibitory effect. The IC₅₀ value of AgNPs was equal to 48.32 µg/mL (42).

In another study, the antioxidant potential of biosynthesized AgNPs using corn leaf waste of *Zea mays* was determined by *in vitro* assays of DPPH radical scavenging against vitamin C as standard. The AgNPs showed a moderate DPPH radical scavenging potential of 34.09% at 100 µg/mL, whereas vitamin C as the reference standard showed higher DPPH scavenging activity of 42.41% at 100 µg/mL (46). However, results for biosynthesized AgNPs are satisfactory.

Anticancer Activity

Sathishkumar et al., proposed the biosynthesis of AgNPs using an aqueous leaf extract of *Alternanthera tenella*. The phytochemical monitoring results showed that flavonoids act as inhibitory and capping agents. The average size of the nanoparticles was found to be 48 nm. The energy-dispersive X-ray spectroscopy (EDX) results demonstrate the AgNPs formation with average size of 40 nm. The

biosynthesized AgNPs are used for the treatment of Human breast adenocarcinoma (MCF7) cells. The IC₅₀ value of the AgNPs was calculated to be 42.5 µg/mL. The AgNPs showed a significant reduction in the migration of MCF-7 cells (34).

In another study, the HCT116 cell lines were treated with different concentrations of biosynthesized AgNPs (50, 100,150, 200, 250, 300 and 350 µg/L) using *Actinidia deliciosa* fruit extract. It is believed that intracellular ROS generation could enhance the anti-cancer activity via nanoparticles. They are dose dependent. The AgNPs treated HCT116 cells showed 78% viability at highest concentration (350 µg/mL) which confirmed the anti-cancer activity of biosynthesized AgNPs (40).

MTT assay results indicated that *Mentha arvensis*-mediated AgNPs could have remarkable cytotoxicity in breast cancer cells (MCF7 and MDA-MB-231). Cell cycle analyses of MCF7 cells exhibited a considerable rise in sub-G1 cell population, confirming the cytotoxicity of AgNPs. On the other hand, human peripheral blood lymphocytes exhibited noticeably less cytotoxicity compared with MCF7 and MDA-MB-231 cells when treated with the same dose. Expression patterns of proteins revealed that AgNPs caused caspase 9-dependent cell death in both cell lines. The Ames test indicated that AgNPs were nonmutagenic in nature (41).

In another study AgNPs were synthesized using *Anthemis atropatana* extract to evaluate their antimicrobial and cytotoxic impacts. The biosynthesized AgNPs have spherical shapes with an average size of 38.89 nm. The MTT results demonstrate the dose dependence of cytotoxic impacts of biosynthesized AgNPs on colon cancer cell lines (HT29). The maximum cytotoxicity impact for biosynthesized AgNPs on HT29 cancer cell line was obtained at 100 µg/mL concentration which was remarkable in terms of statistics when compared with control cells (p<0.001). Moreover, real time PCR and flow cytometry results approved the apoptotic impacts of AgNPs. The results suggest that the green synthesis of AgNPs is an eco-friendly and cost effective approach which could lead to the development of new anticancer and antibacterial agents (44).

Rajakumar et al. examined four different concentrations including; 11.11, 33.33, 100 and 300

Table 1: Properties and applications of biosynthesized AgNPs using plant extract.

Plant	Plant part	Size (nm)	Shape	Application	Ref.
Euphorbia prostrata	Leaves	10-15	Spherical	Leishmanicidal	(24)
Red ginseng	Root	10-30	Spherical	Antibacterial	(25)
Azadirachta indica	Leaves	41-60	Spherical	Biolarvicidal	(26)
Nigella sativa	Leaves	15	Spherical	Cytotoxicity	(27)
Pistacia atlantica	Seeds	27	Spherical	Antibacterial	(28)
Anogeissus latifolia	Gum powder	5.5-5.9	Spherical	Antibacterial	(29)
Tagetes erecta	Flower broth	10-90	Spherical, hexagonal and irregular	Antibacterial and antifungal	(30)
Plumeria rubra	Latex	32-220	Spherical	Mosquito larvicides	(31)
Murraya koenigii	Leaf	20-35	Cubic and spherical	Mosquito larvicides	(32)
Citrullus colocynthis	Calli cells	31	Spherical	Anticancer	(33)
Alternanthera tenella	Leaf	48	Spherical	Anticancer	(34)
Mimosa pudica	Leaf	25-60	Spherical	Antiparasitic	(35)
Olea europaea	Leaves	90	Spherical	Anticancer	(36)
Oak	Fruit hull	40	Spherical	Anticancer	(37)
Solanum tuberosum	Fruit	10	Spherical	Interaction with HSA ^a	(38)
Rhodomyrtus tomentosa	Leaves	10-100	Spherical	antibacterial	(39)
Actinidia deliciosa	Fruit	25-40	Spherical	antioxidant, anticancer and bactericidal	(40)
Mentha arvensis	Leaf	2.8-9.9	Spherical	Anticancer	(41)
Anredera cordifolia	Leaf	40-60	-	Antioxidant and antibacterial	(42)
Tamarindus indica	Fruit	10	Spherical	Antibacterial	(43)
Anthemis atropatana		39	Spherical	Anticancer	(44)
Ipomoea nil	Leaf	-	-	Antibacterial	(45)
Zea mays	Corn leaf waste	45	-	Antioxidant and antibacterial	(46)
Syzygium aromaticum	Flower	5-40	Spherical	Anticancer	(47)
Andrographis paniculata	Leaves	60-70	Hexagonal and spherical	Anticancer	(48)
Milletia pinnata	Flower	49	Spherical	Antibacterial and cytotoxic	(49)
Dunaliella salina	-	15	Spherical	Anticancer	(50)

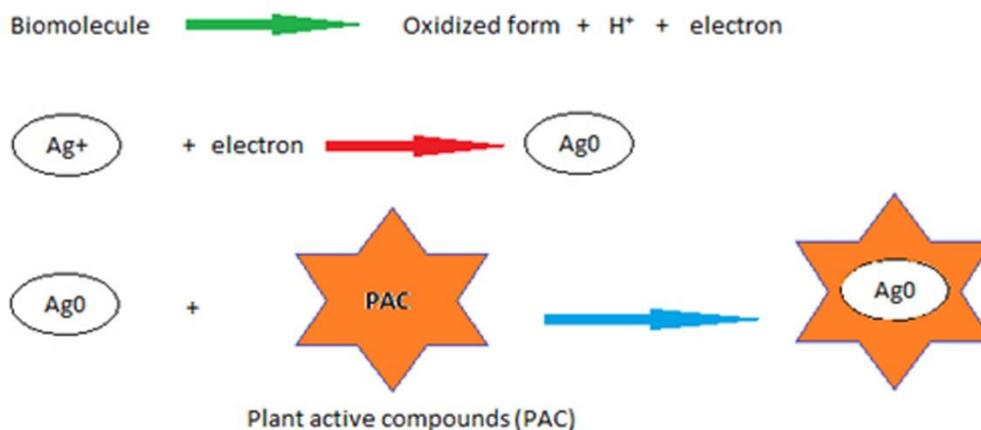
HSA; human serum albumin

mg/mL of synthesized AgNPs and *Millettia pinnata* flower extract to find their cytotoxicity using brine shrimp (*artimia salina*) lethality assay. LD₅₀ value of *M. pinnata* aqueous flower extract was found up to 84.32 mg/mL. The low LD₅₀ value brine shrimp (36.41 mg/mL) exhibited the higher cytotoxic effect of AgNPs compared to *M. pinnata* extract. Cytotoxic effects of AgNPs on shrimp's larvae could be linked with anticancer activity and result in the introduction of an alternative source of anticancer drugs (49).

Anti-inflammatory Capacity

Steroids and non-steroidal anti-inflammatory drugs (NSAIDs), as the main therapeutic agents in inflammation, have serious side effects. Therefore, it is essential to develop new drugs likely to have promising results without serious side effects. Likewise, the anti-inflammatory properties of AgNPs using European black elderberry fruits extract were investigated as in vitro on HaCaT cells exposed to UVB radiation and in vivo on acute inflammation

model and in humans on psoriasis lesions. The results indicated that the biosynthesized AgNPs had a significant anti-inflammatory impact both in vitro and in vivo. In the case of in vitro, the anti-inflammatory effect was confirmed by the reduction of cytokines production and also by keeping their low level after UVB irradiation. In the case of in vivo, the pre-administration of AgNPs led to decline of the level of cytokines in the paw tissues. In the comparison with hydrocortisone, the local treatment of psoriasis vulgaris skin lesions confirmed the significant anti-inflammatory impact of biosynthesized AgNPs (54). The production of ROS, such as superoxide anion, is one of the key factors in inflammation mediated cell damage. In order to examine whether ethanolic petals extract of *Rosa indica* and synthesized AgNPs could have inhibitory effect against H₂O₂ stimulated superoxide anion generation, macrophages were pre-treated with either ethanolic petals extract of *R. indica* and biosynthesized AgNPs. Superoxide anion



Scheme 1. The proposed mechanism for biosynthesis of AgNPs using plant extract.

generation in untreated control was observed, which indicates H₂O₂ stimulated ROS generation in rat peritoneal macrophages. Pre-treatment with both ethanolic extract and synthesized AgNPs significantly inhibited superoxide anion generation in macrophages. The significant reduction was observed in the generation of strong inflammatory mediators such as nitric oxide and superoxide anion upon exposure to synthesized AgNPs which confirm the anti-inflammatory activity of biosynthesized AgNPs (55).

Hepatoprotective Activity of Biosynthesized AgNPs

The hepatoprotective activity of biosynthesized AgNPs using aqueous extracts of *Andrographis paniculata* leaves and *Semecarpus anacardium* nuts against diethylnitrosamine (DEN) induced liver cancer in mice model was investigated. The results of this study revealed that end-capped biomolecules on AgNPs had a potential hepatoprotective impact against DEN induced liver cancer and could be utilized as an efficient anticancer nanodrug (48).

Antilarvicidal Activity

Mosquitoes are capable of transmitting acute human diseases resulting in millions of deaths every year. Synthetic insecticides were utilized to control vector mosquitoes but the result was physiological resistance and pernicious environmental impacts. Furthermore, these synthetic insecticides have high operational cost. Consequently, it is extremely

necessary to introduce insecticides with natural sources for vector control.

The results exhibited by the study by Poopathi et al. indicated that biosynthesized AgNPs that use *Azadirachta indica* have significant larval control over *Aedes aegypti* and *Culex quinquefasciatus*. In this study, the highest effective mortality at LC₅₀ and LC₉₀ levels was observed in *A. aegypti* larvae treated with very low dosage of neem-based AgNPs (LC₅₀ and LC₉₀ equal to 0.006 and 0.04 mg/L, respectively) (26). Moreover, larvicidal activity of synthesized AgNPs using an aqueous extract from *Eclipta prostrate* was observed in crude aqueous and synthesized AgNPs against *Cx. quinquefasciatus* (LC₅₀ = 27.10 and 4.56 mg/L; LC₉₀ = 70.389 and 13.14 mg/L) and *Anopheles subpictus* (LC₅₀ = 27.85 and 5.14 mg/L; LC₉₀ = 71.45 and 25.68 mg/L), respectively (56). In this study, LC₅₀ and LC₉₀ values for biosynthesized AgNPs were higher than the previous report (26). This phenomenon can be attributed to the distinction in end-capping molecules in two AgNPs.

In another research, the activity of biosynthesized AgNPs using *Plumeria rubra* plant latex against *Aedes aegypti* and *Anopheles stephensi* was evaluated. The results indicated that the biosynthesized AgNPs from *P. rubra* latex were much more toxic compared with crude latex extract in both mosquito species. The LC₅₀ values for biosynthesized AgNPs after 24 h of exposure were 1.49, 1.82 mg/L against *A. aegypti* and 1.10, 1.74 mg/L against *A. stephensi*, respectively. On

the other hand, these values for crude latex extract were 181.67, 287.49 mg/L against *A. aegypti* and 143.69, 170.58 mg/L against *A. stephensi*, respectively. This study was the first report on mosquito larvicidal activity of latex synthesized AgNPs (31).

The activity of biosynthesized AgNPs using *Murraya koenigii* plant leaf extract against the first to the fourth instars larvae and pupae of *A. stephensi* and *A. aegypti* was determined. The results indicated that biosynthesized AgNPs from *M. koenigii* leaf were much more toxic compared with crude leaf ethanol extract in both mosquito species. Larvae were exposed to different concentrations of biosynthesized AgNPs and ethanol leaf extract for 24 h. The LC₅₀ values for biosynthesized AgNPs and ethanol leaf extract of *M. koenigii* against *A. stephensi* were 10.82 and 279.33 mg/L, respectively. Moreover, LC₅₀ values for biosynthesized AgNPs and ethanol leaf extract of *M. koenigii* against *A. aegypti* were 13.34 and 314.29 mg/L, respectively. These results demonstrate that biosynthesized AgNPs using *M. koenigii* can be used as a rapid and eco-friendly bio-pesticide which can be developed as a novel approach to produce effective biocides for controlling the target vector mosquitoes (32).

Conclusion

Literature review indicated that the shape and size of biosynthesized AgNPs changed with the variation of extract compounds which could influence the biological applications of these NPs. Plants are rich in medicinally important bio-molecules which act as capping and inhibitory agents for the biosynthesis of AgNPs. The use of plant extract for the synthesis of AgNPs is simple, cost-effective, green and safe. Unlike the chemically synthesized AgNPs, plant-mediated AgNPs are more stable and suitable for biological applications. Biosynthesized AgNPs have significant biological applications, hence, they could contribute to the development of novel drugs for various diseases.

Conflict of Interest

The authors declare that they have no conflict of interest.

References

1. Altavilla C, Ciliberto E. Inorganic Nanoparticles: Synthesis, Applications, and Perspectives, CRC Press, Taylor & Francis Group, 2011, USA.
2. Kesava Kumar CM, Yugandhar P, Savithamma N. Biological synthesis of silver nanoparticles from *Adansonia digitata* L. fruit pulp extract, characterization, and its antimicrobial properties. *J Intercult Ethnopharmacol.* 2016;5:79-85.
3. Yugandhar P, Savithamma N. Leaf assisted green synthesis of silver nanoparticles from *Syzygium alternifolium* Wt. Walp. Characterization and antimicrobial studies. *Nano Biomed Eng.* 2015;7:29-37.
4. Seema G, Amrish C, Avijit M, Rupa M. Green synthesis of silver nanoparticles using *Arnebia nobilis* root extract and wound healing potential of its hydrogel. *Asian J Pharm.* 2014;8:95-101.
5. Sundaravadivelan C, Nalini Padmanabhan M, Sivaprasath P, Kishmu L. Biosynthesized silver nanoparticles from *Pedilanthus tithymaloides* leaf extract with anti-developmental activity against larval instars of *Aedes aegypti* L. Diptera; Culicidae. *Parasitol Res.* 2013;112:303-11.
6. Swamy MK, Sudipta KM, Jayanta K, Balasubramanya S. The green synthesis, characterization, and evaluation of the biological activities of silver nanoparticles synthesized from *Leptadenia reticulata* leaf extract. *Appl Nanosci.* 2014;5:73-81.
7. Vasanth K, Ilango K, Mohan Kumar R, Agrawal A, Dubey GP. Anticancer activity of *Moringa oleifera* mediated silver nanoparticles on human cervical carcinoma cells by apoptosis induction. *Colloids Surf B.* 2014;117:354-9.
8. Rafie HM, Hamed MA. Antioxidant and anti-inflammatory activities of silver nanoparticles biosynthesized from aqueous leaves extracts of four *Terminalia* species. *Adv Nat Sci Nanosci Nanotechnol.* 2014;5:1-11.
9. Bhuvaneshwari R, Chidambaranathan N, Jegatheesan K. Hepatoprotective effect of *Embilica officinalis* and its silver nanoparticles against ccl4 induced hepatotoxicity in wistar albino rats. *Dig J Nanomater Biostruct.* 2014;9:223-35.
10. Kaler A, Mittal AK, Katariya M, Harde H, Agrawal AK, Jain S, et al. An investigation of in vivo wound healing activity of biologically synthesized silver nanoparticles. *J Nanopart Res.* 2014;16:2605
11. Hu R, Yong KT, Roy I, Ding H, He S, Prasad PN. Metallic nanostructures as localized plasmon resonance enhanced scattering probes for multiplex dark field targeted imaging of cancer cells. *J Phys Chem C.* 2009;113:2676-84.
12. Yin B, Ma H, Wang S, Chen S. Electrochemical synthesis of silver nanoparticles under protection of polyN-vinylpyrrolidone. *J Phys Chem B.* 2003;107:8898-904.
13. Dimitrijevic NM, Bartels DM, Jonah CD, Takahashi K, Rajh T. Radiolytically induced formation and optical absorption spectra of colloidal silver nanoparticles in supercritical ethane. *J Phys Chem B* 2001;105:954-9.
14. Wang S, Zhang Y, Ma HL, Zhang Q, Xu W, Peng J, et al. Ionic-liquid-assisted facile synthesis of silver nanoparticle-reduced graphene oxide hybrids by gamma irradiation. *Carbon* 2013;55:245-52.
15. Callegari A, Tonti D, Chergui M. Photochemically grown silver nanoparticles with wavelength-controlled size and shape. *Nano Lett.* 2003;3:1565-8.
16. Pandey S, Goswami GK, Nanda KK. Green synthesis of biopolymer-silver nanoparticle nanocomposite: An optical sensor for ammonia detection. *Int J Biol Macromolec.* 2012;51:583-9.
17. Gopinathan P, Ashok AM, Selvakumar R. Bacterial flagella as biotemplate for the synthesis of silver nanoparticle impregnated bionanomaterial. *Appl Surf Sci.* 2013;276:717-22.
18. Marimuthu S, Abdul Rahuman A, Rajakumar G,

- Santhoshkumar T, Kirthi AV, Jayaseelan C, et al. Evaluation of green synthesized silver nanoparticles against parasites. *Parasitol Res.* 2011;108:1541–9.
19. Nayak RR, Pradhan N, Behera D, Pradhan KM, Mishra S, Sukla LB, et al. Green synthesis of silver nanoparticle by *Penicillium purpurogenum* NPMF: the process and optimization. *J Nanopart Res.* 2011;13:3129–37.
20. Ahmad N, Sharma S. Green synthesis of silver nanoparticles using extracts of *Ananas comosus*. *Green Sustain Chem.* 2012;2:141–7.
21. Sivaraman SK, Elango I, Kumar S, Santhanam V. A green protocol for room temperature synthesis of silver nanoparticles in seconds. *Curr Sci.* 2009;97:1055–59.
22. Martínez-Castañón GA, Niño-Martínez N, Martínez-Gutiérrez F, Martínez-Mendoza JR. Synthesis and antibacterial activity of silver nanoparticles with different sizes. *J Nanopart Res.* 2008;10:1343–8.
23. Lok CN, Ho CM, Chen R, He QY, Yu WY, Sun H, et al. Silver nanoparticles: partial oxidation and antibacterial activities. *J Biol Inorg Chem.* 2007;12:527–34.
24. Zahir AA, Chauhan IS, Bagavan A, Kamaraj C, Elango G, Shankar J, et al. Green Synthesis of Silver and Titanium Dioxide Nanoparticles Using *Euphorbia prostrata* Extract Shows Shift from Apoptosis to G0/G1 Arrest followed by Necrotic Cell Death in *Leishmania donovani*. *Antimicrob Agents Chemother.* 2015;59:4782–99.
25. Singh P, Kim YJ, Wang C, Mathiyalagan R, El-Agamy Farh M, Yang DC. Biogenic silver and gold nanoparticles synthesized using red ginseng root extract, and their applications. *Artif Cells Nanomed Biotechnol.* 2016;44:811–6.
26. Poopathi S, De Britto LJ, Praba VL, Mani C, Praveen M. Synthesis of silver nanoparticles from *Azadirachta indica*—a most effective method for mosquito control. *Environ Sci Pollut Res.* 2015;22:2956–63.
27. Amooghaie R, Saeri MR, Azizi M. Synthesis, characterization and biocompatibility of silver nanoparticles synthesized from *Nigella sativa* leaf extract in comparison with chemical silver nanoparticles. *Ecotoxicol Environ Saf.* 2015;120:400–8.
28. Sadeghi B, Rostami A, Momeni SS. Facile green synthesis of silver nanoparticles using seed aqueous extract of *Pistacia atlantica* and its antibacterial activity. *Spectrochim Acta A Mol Biomol Spectrosc.* 2015;134:326–32.
29. Kora AJ, Beedu SR, Jayaraman A. Size-controlled green synthesis of silver nanoparticles mediated by gum ghatti *Anogeissus latifolia*. and its biological activity. *Org Med Chem Lett.* 2012;2:17.
30. Padalia H, Moteriya P, Chanda S. Green synthesis of silver nanoparticles from marigold flower and its synergistic antimicrobial potential. *Arab J Chem.* 2015;8:732–41.
31. Patil CD, Patil SV, Borase HP, Salunke BK, Salunke RB. Larvicidal activity of silver nanoparticles synthesized using *Plumeria rubra* plant latex against *Aedes aegypti* and *Anopheles stephensi*. *Parasitol Res.* 2012;110:1815–22.
32. Suganya A, Murugan K, Kovendan K, Kumar PM, Hwang JS. Green synthesis of silver nanoparticles using *Murraya koenigii* leaf extract against *Anopheles stephensi* and *Aedes aegypti*. *Parasitol Res.* 2013;112:1385–97.
33. Satyavani K, Gurudeeban S, Ramanathan T, Balasubramanian T. Biomedical potential of silver nanoparticles synthesized from calli cells of *Citrullus colocynthis* (L.) Schrad. *J Nanobiotechnology.* 2011;9:43
34. Sathishkumar P, Vennila K, Jayakumar R, Yusoff AR, Hadibarata T, Palvannan T. Phyto-synthesis of silver nanoparticles using *Alternanthera tenella* leaf extract: an effective inhibitor for the migration of human breast adenocarcinoma MCF-7 cells. *Bioprocess Biosyst Eng.* 2016;39:651–9.
35. Marimuthu S, Rahuman AA, Rajakumar G, Santhoshkumar T, Kirthi AV, Jayaseelan C, et al. Evaluation of green synthesized silver nanoparticles against parasites. *Parasitol Res.* 2011;108:1541–9.
36. Rashidipour M, Heydari R. Biosynthesis of silver nanoparticles using extract of olive leaf: synthesis and in vitro cytotoxic effect on MCF-7 cells. *J Nanostruct Chem.* 2014;4:112
37. Heydari R, Rashidipour M. Green Synthesis of Silver Nanoparticles Using Extract of Oak Fruit Hull Jaft.: Synthesis and in vitro cytotoxic effect on MCF-7 cells. *Int J Breast Cancer.* 2015;Article ID 846743
38. Ali MS, Altaf M, Al-Lohedan HA. Green synthesis of biogenic silver nanoparticles using *Solanum tuberosum* extract and their interaction with human serum albumin: Evidence of “corona” formation through a multi-spectroscopic and molecular docking analysis. *J Photochem Photobiol B.* 2017;173:108–19.
39. Shankar S, Leejae S, Jaiswal L, Voravuthikunchai SP. Metallic nanoparticles augmented the antibacterial potency of *Rhodomlytus tomentosus* acetone extract against *Escherichia coli*. *Microb Pathog.* 2017;107:181–4.
40. Naraginti S, Li Y. Preliminary investigation of catalytic, antioxidant, anticancer and bactericidal activity of green synthesized silver and gold nanoparticles using *Actinidia deliciosa*. *J Photochem Photobiol B.* 2017;170:225–34.
41. Banerjee PP, Bandyopadhyay A, Harsha SN, Policegoudra RS, Bhattacharya S, Karak N, Chattopadhyay A. Mentha arvensis Linn.-mediated green silver nanoparticles trigger caspase 9-dependent cell death in MCF7 and MDA-MB-231 cells. *Breast Cancer: Targets and Therapy.* 2017;9:265–78.
42. Rajathi P, Suja S. Biomimetic synthesis, characterization and evaluation of antioxidant, antimicrobial efficacy of silver nanoparticles using *anredera cordifolia* leaf extract. *Asian J Pharm Clin Res.* 2017;10:329–34.
43. Jayaprakash N, Vijaya JJ, Kaviyarasu K, Kombaiiah K, Kennedy LJ, Ramalingam RJ, et al. Green synthesis of Ag nanoparticles using Tamarind fruit extract for the antibacterial studies. *J Photochem Photobiol B.* 2017;169:178–85.
44. Dehghanizade S, Arasteh J, Mirzaie A. Green synthesis of silver nanoparticles using *Anthemis atropatana* extract: characterization and in vitro biological activities. *Artif Cells Nanomed Biotechnol.* 2017, DOI: 10.1080/21691401.2017.1304402
45. Asha S, Asha A, Rajeshkumar S. Evaluation of phytochemical constituents and antimicrobial activity of silver nanoparticle synthesized ipomoea nil against selected pathogens. *Asian J Pharm Clin Res.* 2017;10:183–7.
46. Patra JK, Baek KH. Antibacterial activity and synergistic antibacterial potential of biosynthesized silver nanoparticles against foodborne pathogenic bacteria along with its anticandidal and antioxidant effects. *Front Microbiol.* 2017;8:167.
47. Venugopal K, Rather HA, Rajagopal K, Shanthi MP, Sheriff K, Illiyas M, et al. Synthesis of silver nanoparticles Ag NPs for anticancer activities MCF 7 breast and A549 lung cell lines of the crude extract of *Syzygium aromaticum*. *J Photochem Photobiol B.* 2017;167:282–9.
48. Prasannaraj G, Venkatachalam P. Hepatoprotective effect of engineered silver nanoparticles coated bioactive compounds against diethylnitrosamine induced hepatocarcinogenesis in experimental mice. *J Photochem Photobiol B.* 2017;167:309–20.
49. Rajakumar G, Gomathi T, Thiruvengadam M, Rajeswari VD, Kalpana VN, Chung M. Evaluation of anti-cholinesterase, antibacterial and cytotoxic activities of green synthesized silver nanoparticles using from *Milletia pinnata* flower extract. *Microb Pathog.* 2017;103:123–8.
50. Singh AK, Tiwari R, Kumar V, Singh P, Riyazat Khadim SK, Tiwari A, et al. Photo-induced biosynthesis of silver nanoparticles from aqueous extract of *Dunaliella salina* and their anticancer

potential. *J Photochem Photobiol B*. 2017;166:202–11.

51. Su HL, Chou CC, Hung DJ, Lin SH, Pao IC, Lin JH, et al. The disruption of bacterial membrane integrity through ROS generation induced by nano hybrids of silver and clay. *Biomater*. 2009;30:5979–87.

52. Gordon O, Slenters TV, Brunetto PS, Villaruz AE, Sturdevant DE, Otto M, et al. Silver coordination polymers for prevention of implant infection: thiol interaction, impact on respiratory chain enzymes, and hydroxyl radical induction. *Antimicrob Agents Chemother*. 2010;54: 4208-18.

53. Hossain Z, Huq F. Studies on the interaction between Cd²⁺ ions and nucleobases and nucleotides. *J Inorgan Biochem*. 2002;90:97-105.

54. David L, Moldovan B, Vulcu A, Olenic L, Perde-Schrepler M, Fischer-Fodor E, et al. Green synthesis, characterization and anti-inflammatory activity of silver nanoparticles using European black elderberry fruits extract. *Colloids Surf B*. 2014;122:767–77.

55. Ramar M, Manikandan B, Raman T, Arunagirinathan K, Prabhu NM, Basu MJ, et al. Biosynthesis of silver nanoparticles using ethanolic petals extract of *Rosa indica* and characterization of its antibacterial, anticancer and anti-inflammatory activities. *Spectrochim Acta A Mol Biomol Spectrosc*. 2015;138:120–9.

56. Rajakumar G, Rahuman AA. Larvicidal activity of silver nanoparticles using *Eclipta prostrata* leaf extract against filariasis and malaria vectors. *Acta Trop*. 2011;118:196–203.